ATTACHMENT A
VERSION WITH MARKINGS TO SHOW CHANGES MADE
(Added language is underlined and deleted language is in brackets)

At page 26, line 9- page 27, line 11, sequence identifiers are assigned to the primer sequences as follows:

Sequences of the primers used for differential display were as follows (purchased from M/s. GenHunter Corporation, USA as a prt of RNA image kit) :

$T_{11}M$ (anchored) primers	**Primer sequence**
$T_{11}A$	5'-AAGCTTTTTTTTTTTTA-3' SEQ ID NO: 5
$T_{11}C$	5'-AAGCTTTTTTTTTTTTC-3' SEQ ID NO: 6
$T_{11}G$	5'-AAGCTTTTTTTTTTTTG-3' SEQ ID NO: 7
Arbitrary Primers	**Primer Sequence**
AP1	5'-AAGCTTGATTGCC-3' SEQ ID NO: 8
AP2	5'-AAGCTTCGACTGT-3' SEQ ID NO: 9
AP3	5'-AAGCTTTGGTCAG-3' SEQ ID NO: 10
AP4	5'-AAGCTTCTCAACG-3' SEQ ID NO: 11
AP5	5'-AAGCTTAGTAGGC-3' SEQ ID NO: 12
AP6	5'-AAGCTTGCACCAT-3' SEQ ID NO: 13
AP7	5'-AAGCTTAACGAGG-3' SEQ ID NO: 14
AP8	5'-AAGCTTTTACCGC-3' SEQ ID NO: 15
AP33	5'-AAGCTTGCTGCTC-3' SEQ ID NO: 16
AP34	5'-AAGCTTCAGCAGC-3' SEQ ID NO: 17
AP35	5'-AAGCTTCAGGGCA-3' SEQ ID NO: 18
AP36	5'-AAGCTTCGACGCT-3' SEQ ID NO: 19
AP37	5'-AAGCTTGGGCCTA-3' SEQ ID NO: 20
AP38	5'-AAGCTTCCAGTGC-3' SEQ ID NO: 21
AP39	5'-AAGCTTTCCCAGC-3' SEQ ID NO: 22

AP40	5'-AAGCTTGTCAGCC-3' SEQ ID NO: 23
AP65	5'-AAGCTT CAAGACC-3' SEQ ID NO: 24
AP66	5'-AAGCTT GCCTTTA-3' SEQ ID NO: 25
AP67	5'-AAGCTT TATTTAT-3' SEQ ID NO: 26
AP68	5'-AAGCTT CTTTGGT-3' SEQ ID NO: 27
AP69	5'-AAGCTT AATAACG-3' SEQ ID NO: 28
AP70	5'-AAGCTT TCATATG-3' SEQ ID NO: 29
AP71	5'-AAGCTT GTAGTAA-3' SEQ ID NO: 30
AP72	5'-AAGCTT TCAAAGA-3' SEQ ID NO: 31

At page 29, the third complete paragraph is amended as follows to assign sequence identifiers to primer sequences:

Single isolated colonies were picked up from re-streaked plates (Example 5) and lysed in 50 µl colony lysis buffer (colony lysis buffer: TE (Tris-Cl 10 mM, 1 mM EDTA, pH 8.0) with 0.1% tween 20) by boiling for 10 minutes. Cell debris were pelleted and the supernatant or the colony lysate containing the template DNA was used for PCR. PCR components were essentially the same as in example 4 except that in place of $T_{11}M$ and arbitrary primers Lgh (5'-CGACAACACCGATAATC-3') SEQ. ID NO: 32 and Rgh (5'-GACGCGAACGAAGCAAC-3') SEQ. ID NO: 33 primers (specific to the vector sequences flanking the cloning site) were used and 2 µl of the colony lysate was used in place of eluted DNA. Also the reaction volume was reduced to 20 µl. PCR conditions used for colony PCR were, 94 °C for 30 seconds, → 52 °C for 40 seconds, → 72 °C for 1 minute for 30 cycles followed by 1 cycle of 5 min extension at 72 °C and final soaking into 4 °C. Amplified product are run on 1.5% agarose gel along with molecular weight marker and analyzed for correct size of insert. While using Lgh and Rgh flanking primers, the size of the cloned PCR product was larger by 120 bp due to the flanking vector sequence being amplified (See Figures 15-17).

At page 32, lines 12-23 were amended as follows to insert sequence identifiers and to correct the number of base pairs at (A).

Each clone was sequenced manually using a T7 sequenase version 2 sequencing kit from M/s. Amersham Pharmacia Biotech, USA. Sequencing primers used were [Lgh (5'-CGACAACACCGATAATC-3') SEQ ID NO: 32 or Rgh (5'-GACGCGAACGAAGCAAC-3') SEQ ID NO: 33]

(1) INFORMATION FOR SEQ ID NO: 1
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30[5]4base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: circular
(ii) MOLECULE TYPE: cDNA
(iii) SEQUENCE DESCRIPTION: SEQ ID NO: 1
Seq ID No. 1# Sequence of 31.2 clone: